

Preliminary Notes

Biogenesis of 6-hydroxylated oestrogens in human tissues

Phenolic steroids hydroxylated at carbon atom 6 have recently gained biological interest. Thus MUELLER AND RUMNEY¹ incubated oestradiol-17 β with mouse liver microsomes in the presence of reduced TPN and obtained evidence for the formation of 6-hydroxyoestradiol-17 β and 6-hydroxyoestrone. BREUER, NOCKE AND KNUPPEN² observed 6-hydroxylation of oestradiol-17 β and oestrone on incubation with rat liver slices, and BREUER AND KNUPPEN³, using the same system, isolated 6-hydroxy-oestradiol-17 β as a metabolite of oestradiol-17 β . Recently MARRIAN⁴ has reported the isolation of a new ketonic Kober chromogen (KC-6B) from pregnancy urine which appears to be identical with a 6-hydroxyoestrone.

There seem to exist two metabolic pathways for the biogenesis of 6-hydroxylated oestrogens in human tissues, one involving the conversion of 6-hydroxylated C₁₉-steroids to the corresponding C₁₈-steroids, and the second which involves the direct hydroxylation of oestradiol-17 β . We have examined both possibilities (1) by using RYAN's enzyme preparation of human placenta⁵ which converts readily Δ^4 -androstene-3,17-dione to oestrone and (2) by carrying out incubations with human fetal liver slices.

Human placentae were prepared as described by RYAN⁵. The 10,000 \times g supernatant fractions each equivalent to 15 g of tissue were incubated with 10 μ moles ATP, 2.5 μ moles DPN and 200 μ g of 6 α - or 6 β -hydroxy- Δ^4 -androstene-3,17-dione at 37° for 60 min in a total volume of 5 ml. The incubation mixtures were extracted with ether-chloroform (3:1, v/v) and the phenolic fractions chromatographed on paper in the formamide-chloroform system. The paper chromatograms were sprayed with FOLIN AND CIOCALTEU reagent^{6,7}. After incubation of 6 α -hydroxy- Δ^4 -androstene-3,17-dione a phenolic metabolite (I) was detected which showed the same mobility (14.2 cm/24 h) as authentic 6' α '-hydroxyoestrone prepared by oxidation with chromic acid from 6' α '-hydroxyoestradiol-17 β 3-monoacetate 6-monobenzoate⁸. (I) was a Kober chromogen and had the same absorption curve as 6' α '-hydroxyoestrone with the H₂SO₄-H₂O reaction (λ_{\max} . 270 m μ , 289 m μ , 462 m μ ; λ_{\min} . 260 m μ , 275 m μ , 360 m μ). For further identification (I) was sublimed under normal pressure. The crystals obtained had a melting point (261–263°) which was identical with that of 6-dehydrooestrone; the same melting point was observed after sublimation of authentic 6' α '-hydroxyoestrone. The phenolic substance (II) detected after incubation of 6 β -hydroxy- Δ^4 -androstene-3,17-dione showed a similar mobility (14.6 cm/24 h) as 6' α '-hydroxyoestrone. On rechromatography in the system benzene-light petroleum-methanol-water (33:66:80:20) the mobility of (II) was slightly greater (7.6 cm/24 h) than that of 6' α '-hydroxyoestrone (6.8 cm/24 h). (II) gave a red colour with the Kober reaction. From the data reported here it is concluded that human

Abbreviations: TPN, triphosphopyridine nucleotide; DPN, diphosphopyridine nucleotide; ATP, adenosine triphosphate.

placenta can convert 6 α -hydroxy- Δ^4 -androstene-3,17-dione to 6 α -hydroxyoestrone and 6 β -hydroxy- Δ^4 -androstene-3,17-dione to 6 β -hydroxyoestrone. It is interesting to note that approx. 4 times as much 6 α -hydroxyoestrone was formed as 6 β -hydroxyoestrone.

In order to investigate the second metabolic pathway leading to the formation of 6-hydroxylated oestrogens, oestradiol-17 β was incubated with human fetal liver. 200 mg of liver slices were shaken with 200 μ g of steroid in 5 ml Krebs-phosphate saline at 37° for 60 min. The incubation mixtures were extracted with chloroform-ether (3:1, v/v) and the extracts evaporated. The residues were subjected to paper chromatography in the system formamide-chloroform. The "polar" fractions containing oestriol and possibly 6' α '-hydroxyoestradiol-17 β were eluted from the paper and methylated with dimethyl sulfate; the methylated material was then applied to an alumina column (cf. ref. 2). Upon elution with 1.4 % ethanol in benzene a Kober chromogen was obtained which showed in the H₂SO₄-H₂O reaction the same absorption curve as the 3-methyl ether of 6' α '-hydroxyoestradiol-17 β (λ_{max} , 288 m μ , 438 m μ , 462 m μ ; λ_{min} , 260 m μ , 350 m μ , 450 m μ). Further elution with 3.0 % ethanol in benzene yielded the 3-methyl ether of oestriol. On the basis of these results it is concluded that oestradiol-17 β is hydroxylated by human fetal liver not only to oestriol—as has already been shown by ENGEL, BAGGETT AND HALLA⁹—but also to 6' α '-hydroxyoestradiol-17 β . It seems therefore that, with respect to the metabolism of oestradiol-17 β , human fetal liver resembles very closely that of rat liver.

Full experimental details will be published elsewhere. The authors wish to express their gratitude to Professor M. EHRENSTEIN, University of Pennsylvania, Philadelphia, for a generous gift of 6 α - and 6 β -hydroxy- Δ^4 -androstene-3,17-dione, and to Professor C. DJERASSI, Wayne State University, Detroit, for a sample of 6-dehydrooestrone. This work was supported by the Deutsche Forschungsgemeinschaft.

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Received March 23rd, 1960